MANUAL

Adiponectin (rhesus monkey, macaque)

Competitive ELISA Kit

For research use only. Not for diagnostic use.

Version 2 (14-March-2011)

Cat. No. AG-45A-0003EK-KI01

www.adipogen.com
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1. Intended Use

The Adiponectin (rhesus monkey, macaque) Competitive ELISA Kit is to be used for the in vitro quantitative determination of monkey adiponectin in serum, plasma and cell culture supernatant. This ELISA Kit is for research use only.

2. Introduction

Adipocytes express a variety of adipocytokines that function in the homeostatic control of glucose and lipid metabolism. Insulin regulates secretion of many of these adipocytokines in response to changes in energy balance. Adiponectin is a 244-amino acid protein with high structural homology to collagen VIII, collagen V, complement C1q(Ref. 1 and 2), and TNF(Ref.3), which is exclusively and abundantly expressed in white adipose tissue. Plasma adiponectin concentrations have found to be decreased in obesity and/or type-2 diabetes, resulting in the conditions commonly associated with insulin resistance and hyper-insulinemia(Ref. 4-5). Therefore, measurement of the plasma level of adiponectin may be important for understanding diagnosis or prognosis of onset of these diseases.
3. General References


(3) The crystal structure of a complement-1q family protein suggests an evolutionary link to tumor necrosis: L. Shapiro, et al.; Curr. Biol. 12, 335 (1998)


(6) Circulating concentrations of adipocyte protein adiponectin are decreased in parallel with reduced insulin sensitivity during the progression to type 2 diabetes in rhesus monkey: K. Hotta, et al.; Diabetes 50, 1126 (2001)
4. Assay Principle

This assay is a competitive Enzyme Linked-Immunosorbent Assay (ELISA) for quantitative determination of monkey adiponectin in biological fluids. A polyclonal antibody recognizing native monkey adiponectin reacts with a series of predetermined recombinant monkey adiponectin standard proteins or samples under competition in the adiponectin-coated plate. Their relative reactivity is plotted with that of the standard proteins.

5. Handling & Storage

- Reagent must be stored at 2-8°C when not in use.
- Plate and reagents should be at room temperature before use.
- Do not expose reagents to temperatures greater than 25°C.

6. Kit Components

1 plate coated with monkey adiponectin Recombinant Protein (12 x 8-well strips)
1 bottle Wash Buffer 10X (50 ml)
1 bottle Diluent 5X (50 ml)
1 bottle Detection Antibody (12 ml)
1 vial Detector 100X (HRP Conjugated anti-rabbit IgG) (150 µl)
1 vial monkey adiponectin Standard (lyophilized) (1 µg)
1 vial monkey adiponectin QC sample (lyophilized)
1 bottle Substrate Solution I (TMB) (6 ml)
1 bottle Substrate Solution II (Peroxidase) (6 ml)
1 bottle Stop Solution (12 ml)
3 plate sealers (plastic film)
7. Materials Required but Not Supplied

- Microtiterplate reader at 450 nm, with the correction wavelength set at 540 nm or 570 nm
- Calibrated precision single and multi-channel pipettes. Disposable pipette tips
- Deionized water
- Microtubes or equivalent for preparing dilutions
- Disposable plastic containers for preparing working buffers
- Plate washer: automated or manual
- Glass or plastic tubes for diluting and aliquoting standard
8. General ELISA Protocol

8.1. Preparation and Storage of Reagents

NOTE: Prepare just the appropriate amount of the buffers necessary for the assay.

- **Wash Buffer 10X** has to be diluted with deionized water 1:10 before use (e.g. 50 ml Wash Buffer 10X + 450 ml water) to obtain Wash Buffer 1X.

- **Diluent 5X** has to be diluted with deionized water 1:5 before use (e.g. 50 ml Diluent 5X + 200 ml water) to obtain Diluent 1X.

- **Detector 100X (HRP Conjugated anti-rabbit IgG)** has to be diluted to the working concentration by adding 120 µl in 12 ml of Diluent 1X (1:100).

  NOTE: The diluted Detector is used within one hour of preparation.

- **Substrate Solution I and II** have to be mixed together in equal volumes within 15 minutes of use.

  NOTE: Freshly prepare just before use the Substrate Solution and protect from light!

- **Monkey adiponectin Standard (STD)** has to be reconstituted with 1 ml of deionized water.

  o This reconstitution produces a stock solution of 1 µg/ml. Mix the standard to ensure complete reconstitution and allow the standard to sit for a minimum of 15 minutes. Mix well prior to making dilutions.

  NOTE: The reconstituted standard is aliquoted and stored at -20°C.

  o Dilute the standard protein concentrate (STD) (1 µg/ml) in Diluent 1X. A seven-point standard curve in Diluent 1X is recommended.

  o Suggested standard points are:

    1, 0.5, 0.25, 0.1, 0.05, 0.025, 0.01 and 0.001 µg/ml.

- **Monkey adiponectin QC sample** has to be reconstituted with 1 ml of deionized water.

  o Refer to the Certificate of Analysis for current QC sample concentration. Mix the QC sample to ensure complete reconstitution and allow the QC sample to sit for a minimum of 15 minutes. The reconstituted QC sample is ready to use, do not dilute it.
Dilute further for the standard curve:

<table>
<thead>
<tr>
<th>To obtain</th>
<th>Add</th>
<th>Into</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 µg/ml</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>0.5 µg/ml</td>
<td>300 µl of adiponectin (1 µg/ml)</td>
<td>300 µl of Diluent 1X</td>
</tr>
<tr>
<td>0.25 µg/ml</td>
<td>300 µl of adiponectin (0.5 µg/ml)</td>
<td>300 µl of Diluent 1X</td>
</tr>
<tr>
<td>0.1 µg/ml</td>
<td>200 µl of adiponectin (0.25 µg/ml)</td>
<td>300 µl of Diluent 1X</td>
</tr>
<tr>
<td>0.05 µg/ml</td>
<td>300 µl of adiponectin (0.1 µg/ml)</td>
<td>300 µl of Diluent 1X</td>
</tr>
<tr>
<td>0.025 µg/ml</td>
<td>300 µl of adiponectin (0.05 µg/ml)</td>
<td>300 µl of Diluent 1X</td>
</tr>
<tr>
<td>0.01 µg/ml</td>
<td>200 µl of adiponectin (0.025 µg/ml)</td>
<td>300 µl of Diluent 1X</td>
</tr>
<tr>
<td>0.001 µg/ml</td>
<td>50 µl of adiponectin (0.01 µg/ml)</td>
<td>450 µl of Diluent 1X</td>
</tr>
</tbody>
</table>

8.2. Sample Collection, Storage and Dilution

**Serum**: Use a serum separator tube. Let samples clot at room temperature for 30 minutes before centrifugation for 20 minutes at 1,000xg. Assay freshly prepared serum or store serum in aliquot at ≤ -20°C for later use. Avoid repeated freeze/thaw cycles.

**Plasma**: Collect plasma using heparin, EDTA, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000xg within 30 minutes of collection. Assay freshly prepared plasma or store plasma sample in aliquot at ≤ -20°C for later use. Avoid repeated freeze/thaw cycles.

**Serum, Plasma or Cell Culture Supernatant** have to be diluted in Diluent 1X. Samples containing visible precipitates must be clarified before use.

**NOTE**: As a starting point, 1/1,000 dilution of serum or plasma is recommended! If samples fall the outside range of assay, a lower or higher dilution may be required!
### 8.3. Assay Procedure (Checklist)

1. Determine the number of 8-well strips needed for the assay and insert them in the frame for current use. The extra strips should be resealed in the foil pouch bag and stored at 4°C.  
   **NOTE:** Remaining 8-well strips coated with adiponectin protein when opened can be stored at 4°C for up to 1 month.

2. Add 50 µl of the different standards and reconstituted QC sample into the appropriate wells in duplicate! At the same time, add 50 µl of diluted serum, plasma or cell culture supernatant samples in duplicate to the wells (see 8.1. Preparation and Storage of Reagents and 8.2. Preparation of Samples).

3. Add 50 µl to each well of the Detection Antibody and tap gently on the side of the plate to mix.

4. Cover the plate with plate sealer and incubate for **2 hours at room temperature (RT°C) on the shaker.**

5. Aspirate the coated wells and add 300 µl of Wash Buffer 1X using a multi-channel pipette or auto-washer. Repeat the process for a total of three washes. After the last wash, complete removal of liquid is essential for good performance.

6. Add 100 µl to each well of the diluted Detector (see 8.1. Preparation and Storage of Reagents).

7. Cover the plate with plate sealer and incubate for **1 hour at RT°C on the shaker.**

8. Aspirate the coated wells and add 300 µl of Wash Buffer 1X using a multi-channel pipette or auto-washer. Repeat the process for a total of five washes. After the last wash, complete removal of liquid is essential for good performance.

9. Add 100 µl to each well of mixed substrate solution.

10. Allow the color reaction to develop at **RT°C in the dark for 20 minutes.**

11. Stop the reaction by adding 100 µl of Stop Solution. Tap the plate gently to ensure thorough mixing. The substrate reaction yields a blue solution that turns yellow when Stop Solution is added. **! CAUTION: CORROSIVE SOLUTION!**

12. Measure the OD at 450 nm in an ELISA reader within 30 minutes.
9. Calculation of Results

- Average the duplicate readings for each standard, QC and sample.
- Generate the standard curve by plotting the average absorbance obtained for each standard concentration on the vertical (Y) axis vs. the corresponding adiponectin concentration (µg/ml) on the horizontal (X) axis (see 10. TYPICAL DATA).
- Calculate the adiponectin concentrations of samples by interpolation of the regression curve formula as shown above in a form of a 4-parameter logistic equation.
- If the test samples were diluted, multiply the interpolated values by the dilution factor to calculate the concentration of monkey adiponectin in the samples.

10. Typical Data

The following data are obtained using the different concentrations of standard as described in this protocol:

<table>
<thead>
<tr>
<th>Standard monkey Adiponectin (µg/ml)</th>
<th>Optical Density (mean)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>0.215</td>
</tr>
<tr>
<td>0.5</td>
<td>0.289</td>
</tr>
<tr>
<td>0.25</td>
<td>0.451</td>
</tr>
<tr>
<td>0.1</td>
<td>0.821</td>
</tr>
<tr>
<td>0.05</td>
<td>1.237</td>
</tr>
<tr>
<td>0.025</td>
<td>1.665</td>
</tr>
<tr>
<td>0.01</td>
<td>2.14</td>
</tr>
<tr>
<td>0.001</td>
<td>2.489</td>
</tr>
</tbody>
</table>

**Figure:** Standard curve
11. Performance Characteristics

A. Sensitivity (Limit of detection):

The lowest level of adiponectin that can be detected by this assay is 1 ng/ml. **NOTE: The Limit of detection was measured by adding two standard deviations to the mean value of 50 zero standard.**

B. Assay range: 0.001 µg/ml – 1 µg/ml

C. Specificity:

This ELISA is specific for the measurement of natural and recombinant monkey adiponectin. It does not cross-react with mouse adiponectin, rat adiponectin, human resistin, human RELM-α, human RELM-β, human leptin, human TNF-α.

Human adiponectin shows 100% cross-reactivity in this assay.

D. Intra-assay precision:

Three samples of known concentrations of monkey adiponectin were assayed in replicates 5 times to test precision within an assay.

<table>
<thead>
<tr>
<th>Samples</th>
<th>Means (µg/ml)</th>
<th>SD</th>
<th>CV (%)</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>16.82</td>
<td>0.44</td>
<td>2.64</td>
<td>5</td>
</tr>
<tr>
<td>2</td>
<td>14.13</td>
<td>1.13</td>
<td>7.98</td>
<td>5</td>
</tr>
<tr>
<td>3</td>
<td>12.55</td>
<td>0.72</td>
<td>5.75</td>
<td>5</td>
</tr>
</tbody>
</table>

E. Inter-assay precision:

Three samples of known concentrations of monkey adiponectin were assayed in 5 separate assays to test precision between assays.

<table>
<thead>
<tr>
<th>Samples</th>
<th>Means (µg/ml)</th>
<th>SD</th>
<th>CV (%)</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>18.66</td>
<td>0.73</td>
<td>3.89</td>
<td>5</td>
</tr>
<tr>
<td>2</td>
<td>16.29</td>
<td>0.90</td>
<td>5.50</td>
<td>5</td>
</tr>
<tr>
<td>3</td>
<td>11.87</td>
<td>0.75</td>
<td>6.33</td>
<td>5</td>
</tr>
</tbody>
</table>
F. Linearity:

Different monkey serum samples containing adiponectin were diluted several fold (1/1,000 to 1/4,000) and the measured recoveries ranged from 80% to 100%.

<table>
<thead>
<tr>
<th>Samples</th>
<th>Sample Dilution</th>
<th>Expected (µg/ml)</th>
<th>Observed (µg/ml)</th>
<th>% of Expected</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1 : 1,000</td>
<td>14</td>
<td>14</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>1 : 2,000</td>
<td>7</td>
<td>6</td>
<td>86</td>
</tr>
<tr>
<td></td>
<td>1 : 4,000</td>
<td>3.5</td>
<td>3</td>
<td>86</td>
</tr>
<tr>
<td>2</td>
<td>1 : 1,000</td>
<td>15</td>
<td>15</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>1 : 2,000</td>
<td>7.5</td>
<td>6</td>
<td>80</td>
</tr>
<tr>
<td></td>
<td>1 : 4,000</td>
<td>3.75</td>
<td>3</td>
<td>80</td>
</tr>
</tbody>
</table>
12. Technical Hints and Limitations

- It is recommended that all standards, QC sample and samples be run in duplicate.
- Do not combine leftover reagents with those reserved for additional wells.
- Reagents from the kit with a volume less than 100 µl should be centrifuged.
- Residual wash liquid should be drained from the wells after last wash by tapping the plate on absorbent paper.
- Crystals could appear in the 10X solution due to high salt concentration in the stock solutions. Crystals are readily dissolved at room temperature or at 37°C before dilution of the buffer solutions.
- Once reagents have been added to the 8-well strips, DO NOT let the strips DRY at any time during the assay.
- Keep Substrate Solution protected from light.
- The Stop Solution consists of phosphoric acid. Although diluted, the Stop Solution should be handled with gloves, eye protection and protective clothing.
13. Troubleshooting

<table>
<thead>
<tr>
<th>PROBLEM</th>
<th>POSSIBLE CAUSES</th>
<th>SOLUTIONS</th>
</tr>
</thead>
<tbody>
<tr>
<td>No signal or weak signal</td>
<td>Omission of key reagent</td>
<td>Check that all reagents have been added in the correct order.</td>
</tr>
<tr>
<td></td>
<td>Washes too stringent</td>
<td>Use an automated plate washer if possible.</td>
</tr>
<tr>
<td></td>
<td>Incubation times inadequate</td>
<td>Incubation times should be followed as indicated in the manual.</td>
</tr>
<tr>
<td></td>
<td>Plate reader settings not optimal</td>
<td>Verify the wavelength and filter setting in the plate reader.</td>
</tr>
<tr>
<td></td>
<td>Incorrect assay temperature</td>
<td>Use recommended incubation temperature. Bring substrates to room temperature before use.</td>
</tr>
<tr>
<td>High background</td>
<td>Concentration of detector too high</td>
<td>Use recommended dilution factor.</td>
</tr>
<tr>
<td></td>
<td>Inadequate washing</td>
<td>Ensure all wells are filling wash buffer and are aspirated completely.</td>
</tr>
<tr>
<td>Poor standard curve</td>
<td>Wells not completely aspirated</td>
<td>Completely aspirate wells between steps.</td>
</tr>
<tr>
<td></td>
<td>Reagents poorly mixed</td>
<td>Be sure that reagents are thoroughly mixed.</td>
</tr>
<tr>
<td>Unexpected results</td>
<td>Omission of reagents</td>
<td>Be sure that reagents were prepared correctly and added in the correct order.</td>
</tr>
<tr>
<td></td>
<td>Dilution error</td>
<td>Check pipetting technique and double-check calculations.</td>
</tr>
</tbody>
</table>
14. Assay Flow Chart

1. Prepare reagents, samples and Standards as instructed.

2. Add 50 µl of Standards, QC and samples to each well.

3. Add 50 µl of Detection Antibody to each well.

4. Incubate for 2 hours at RT°C on the shaker.

5. Aspirate and wash 3 times.

6. Add 100 µl of diluted Detector to each well.

7. Incubate for 1 hour at RT°C on the shaker.

8. Aspirate and wash 5 times.

9. Add 100 µl of mixed Substrate Solution to each well. Incubate for 20 mins at RT°C in the dark.

10. Add 100 µl of Stop Solution to each well. Read at 450 nm within 30 mins.
Product Specific References:

For more References please visit www.adipogen.com!