# **EDI™** Human ACTH ELISA

Enzyme Linked ImmunoSorbent Assay (ELISA) for the Quantitative Measurement of Human Adrenocorticotropic hormone (ACTH) Levels in EDTA-Plasma



KT-882













#### **INTENDED USE**

This test kit is intended for use in the quantitative determination of human adrenocorticotropic hormone (ACTH) in EDTA-plasma. The test is useful for detecting elevated and deficient ACTH levels.

### Indications for use:

Patient may have a higher than normal levels of ACTH with

- Addison's disease or primary adrenal insufficiency
- Congenital adrenal hyperplasia
- 3. Cushing's syndrome
- Cushing's disease 4.
- Multiple endocrine neoplasia (MEN), type I

Patient may have a lower than normal levels of ACTH with

- Hypopituitarism and/or secondary adrenal insufficiency
- 7. Adrenal gland tumor
- Other tumors that produce cortisol

#### SUMMARY OF PHYSIOLOGY

ACTH is a 39 amino acid polypeptide with a molecular weight of 4540 Dalton. ACTH is secreted from corticotropes in the anterior lobe (or adenohypophysis) of the pituitary gland in response to corticotropin-releasing hormone (CRH) released by the hypothalamus. ACTH is synthesized from pre-pro-opiomelanocortin (pre-POMC). The removal of the signal peptide during translation produces the 241-amino acid polypeptide POMC, which undergoes a series of post-translational modifications such as phosphorylation and glycosylation before it is proteolytically cleaved by endopeptidases to yield various polypeptide fragments with varying physiological activity.

ACTH is an important component of the hypothalamic-pituitaryadrenal axis and is often produced in response to biological stress. It stimulates secretion of glucocorticoid steroid hormones from adrenal cortex cells especially in the zona fasciculata of the adrenal. ACTH acts by binding to cell surface ACTH receptors, which are located primarily on adrenocortical cells of the adrenal cortex.

# **ASSAY PRINCIPLE**

This ELISA kit is designed, developed and produced for the quantitative measurement of human ACTH in EDTA-plasma sample. The assay utilizes the two-site "sandwich" technique with selected antibodies that bind to N-terminal and C-terminal epitopes of ACTH.

Assay standards, controls and patient samples are added directly to wells of a microtiter plate that is coated with antibody to the Cterminal of human ACTH. Immediately, a horseradish peroxidase (HRP) conjugated anti N-terminal of human ACTH antibody is added to each well. After the first incubation period, a "sandwich" of solidphase polyclonal antibody - human ACTH - HRP conjugated monoclonal antibody" is formed. The unbound antibodies and buffer matrix are removed in the subsequent washing step. For the detection of this immunocomplex, the well is then incubated with a

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substrate solution in a timed reaction, which is terminated with an acidic reagent (i.e. ELISA stop solution). The absorbance is then measured in a spectrophotometric microplate reader. The enzymatic activity of the immunocomplex bound to the wall of each microtiter well is directly proportional to the amount of human ACTH in the test sample. A standard curve is generated by plotting the absorbance versus the respective human ACTH concentration for each standard on a point-to-point or 4-parameter curve fitting. The concentration of human ACTH in test samples is determined directly from this standard curve.

#### **REAGENTS: Preparation and Storage**

This test kit must be stored at 2 - 8°C upon receipt. For the expiration date of the kit refer to the label on the kit box. All components are stable until this expiration date.

Prior to use allow all reagents to come to room temperature. Regents from different kit lot numbers should not be combined or interchanged.

# Anti-ACTH Antibody Coated Microplate (Cat. No.

One microplate with twelve by eight strips (96 wells total) coated with polyclonal anti-human ACTH antibody. The plate is framed and sealed in a foil zipper bag with a desiccant. This reagent should be stored at 2 - 8°C and is stable until the expiration date on the kit box.

HRP Conjugated Anti-ACTH Antibody (Cat. No. 30639) One vial containing 0.25 mL HRP-labeled anti-human ACTH antibody in a stabilized protein matrix. This reagent should be stored at 2 - 8°C and is stable until the expiration date on the kit box.

#### ELISA Wash Concentrate (Cat. No. 10010)

One bottle containing 30 mL of 30-fold concentrate. Before use the contents must be diluted with 870 mL of demineralized water and mixed well. Upon dilution, this yields a working wash solution containing a surfactant in phosphate-buffered saline with a non-azide, non-mercury preservative. The diluted wash solution may be stored at room temperature and is stable until the expiration date on the kit box.

#### ELISA HRP Substrate (Cat. No. 10020)

One bottle containing 25 mL of tetramethylbenzidine (TMB) with hydrogen peroxide. This reagent should be stored at 2 - 8°C and is stable until the expiration date on the kit box.

#### ELISA Stop Solution (Cat. No. 30357)

One bottle containing 12 mL of stop solution. This reagent may be stored at 2 – 8°C or room temperature and is stable until the expiration date on the kit box.

#### 6. Human ACTH Standards (Cat. No. 30631-30636)

Six vials containing human ACTH in a lyophilized bovine serumbased matrix with a non-azide preservative. **Refer to the vials for exact concentration of the standard.** These standards should be stored at 2 – 8°C and are stable until the expiration date on the kit box. Refer to assay procedure section for dilution direction.

#### 7. Human ACTH Controls (Cat. No. 30637 - 30638)

Two vials containing human ACTH in a lyophilized bovine serum-based matrix with a non-azide preservative. **Refer to vials for exact concentration range for each control.** Both controls should be stored at 2 – 8°C and are stable until the expiration date on the kit box. Refer to assay procedure section for reconstitution instructions.

#### 8. Tracer Antibody Diluent (Cat. No. 30640)

One vial containing 5 mL ready-to-use buffer. It should be used only for tracer antibody dilution according to the assay procedures. This reagent should be stored at  $2-8^{\circ}$ C and is stable until the expiration date on the kit box.

#### SAFETY PRECAUTIONS

The reagents must be used in professional laboratory. Source material for reagents containing bovine serum was derived in the contiguous 48 United States. It was obtained only from healthy donor animals maintained under veterinary supervision and found free of contagious diseases. Wear gloves while performing this assay and handle these reagents as if they are potentially infectious. Avoid contact with reagents containing TMB, hydrogen peroxide, or sulfuric acid. TMB may cause irritation to skin and mucous membranes and cause an allergic skin reaction. TMB is a suspected carcinogen. Sulfuric acid may cause severe irritation on contact with skin. Do not get in eyes, on skin, or on clothing. Do not ingest or inhale fumes. On contact, flush with copious amounts of water for at least 15 minutes. Use Good Laboratory Practices.

#### MATERIALS REQUIRED BUT NOT PROVIDED

- Precision single channel pipettes capable of delivering 25 μL, 200 μL, etc.
- Disposable pipette tips suitable for above volume dispensing.
- Aluminum foil.
- 4. Deionized or distilled water.
- Plastic microtiter well cover or polyethylene film.
- ELISA multichannel wash bottle or automatic (semiautomatic) washing system.
- Spectrophotometric microplate reader capable of reading absorbance at 450/650 or 450/620 nm.

#### **SPECIMEN COLLECTION**

Since the circulating ACTH shows a 24 hours circadian rhythms, it is recommended to draw blood sample in the early morning, before 8 a.m. Patients should stop taking steroid drugs before drawing blood sample, at the consultation of their physician.

EDTA-plasma is a suitable specimen for human ACTH measurement. A total of 0.4 mL EDTA-plasma is required for duplicate determination of human ACTH with this test kit. Whole blood should be collected using lavender-top Vaccutainer and the plasma separated according to manufacturer's instruction. The EDTA-plasma should be separated from other cells right after or within one hour of blood collection. The plasma should be transferred to a clean test tube right after centrifugation. **Plasma samples should be stored at – 20°C** if the assay is not to be performed within 3 hours. Avoid more than three times freeze-thaw cycles of specimen.

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Samples of serum, heparin plasma and citrate plasma should not be used for ACTH measurement.

#### **ASSAY PROCEDURE**

#### 1. Reagent Preparation

- (1) Prior to use allow all reagents to come to room temperature. Reagents from different kit lot numbers should not be combined or interchanged.
- (2) ELISA Wash Concentrate (Cat. 10010) must be diluted to working solution prior to use. Please see REAGENTS section for details.
- (3) Reconstitute assay standards and controls by adding 2.0 mL of deminerialized water to each standard and control bottle. Allow the standard and controls to sit undisturbed for 5 minutes, and then mix well by inversions or gentle vortexing. One must make sure that all solid is dissolved completely prior to use. These reconstituted standards and controls may be stored at 2- 8°C for up to 24 hours or below -10 °C for long-term storage. Do not exceed 3 freeze-thaw cycles.
- (4) Prepare Tracer Antibody working solution by 1:21 fold dilution of the ACTH Tracer Antibody (Cat. 30639) by adding the tracer antibody into the Tracer Antibody Diluent (Cat. 30640). Following is a table that outlines the relationship of strips used and antibody mixture prepared. NOTE: the tracer antibody should be prepared just prior to the beginning of the assay.

Dilution Scheme	Tracer Antibody Diluent	Tracer Antibody
1	0.4 mL	20 μL
2	0.8 mL	40 μL
3	1.2 mL	60 µL
4	1.6 mL	80 µL
5	2.0 mL	100 μL
6	2.4 mL	120 µL
7	2.8 mL	140 µL
8	3.2 mL	160 µL
9	3.6 mL	180 µL
10	4.0 mL	200 μL
11	4.4 mL	220 µL
12	4.8 mL	240 µL

#### (5) Test Configuration

ROW	STRIP 1	STRIP 2	STRIP 3	STRIP 4
Α	STD 1	STD 5	SAMPLE 1	SAMPLE 5
В	STD 1	STD 5	SAMPLE 1	SAMPLE 5
С	STD 2	STD 6	SAMPLE 2	SAMPLE 6
D	STD 2	STD 6	SAMPLE 2	SAMPLE 6
E	STD 3	C 1	SAMPLE 3	
F	STD 3	C 1	SAMPLE 3	
G	STD 4	C 2	SAMPLE 4	
Н	STD 4	C 2	SAMPLE 4	

(6) Place a sufficient number of Anti-ACTH antibody-coated microwell strips in a holder to run human ACTH standards, controls and unknown samples in duplicates.

#### 2. Assay Procedure:

- (1) Add **200 µL** of standards, controls and patient samples into the designated microwells.
- (2) Immediately add 25 μL of HRP Conjugated Anti-ACTH Tracer Antibody mix to each well.
- (3) Seal the plate wells securely, cover with foil or other material to protect from light, and rotate on an ELISA plate shaker (small orbit radius) for 2 hr. ± 5 minutes at 400 to 450 rpm.
- (4) Wash each well 5 times by dispensing 350 µL of working wash solution into each well and then completely aspirating the contents. Alternatively, an automated microplate washer can be used.
- (5) Add 200 μL of ELISA HRP Substrate (Cat. 10020) into each of the wells.
- (6) Cover the plate with aluminum foil or other material to avoid exposure to light. Incubate plate static, at room temperature for 20 minutes.
- (7) Immediately add **50 µL** of ELISA Stop Solution (Cat. 30557) into each of the wells. Mix gently.
- (8) Read the absorbance at 450 nm with reference filter at 620 nm or 650 nm.

#### **PROCEDURAL NOTES**

- It is recommended that all standards, controls and unknown samples be assayed in duplicate. The average absorbance reading of each duplicate should be used for data reduction and the calculation of results.
- 2. Keep light-sensitive reagents in the original amber bottles.
- Store any unused antibody-coated strips in the foil zipper bag with desiccant to protect from moisture.
- Careful technique and use of properly calibrated pipetting devices are necessary to ensure reproducibility of the test.
- 5. Incubation times or temperatures other than those stated in this insert may affect the results.
- An orbital mixer with a larger orbit radius (e.g. > 1 cm) may be used at speeds of 150 to 200 rpm.
- Avoid air bubbles in the microwell as this could result in lower binding efficiency and higher CV% of duplicate reading.
- All reagents should be mixed gently and thoroughly prior to use. Avoid foaming.
- If adapting this assay to automated ELISA system such as DS-2 (Diamedix Corp., Miami), a procedural validation is necessary if there is any modification of the assay procedure.

#### INTERPRETATION OF RESULTS

It is recommended to use a point-to-point or 4-parameter standard curve fitting.

- Calculate the average absorbance for each pair of duplicate test results.
- Subtract the average absorbance of the level 1 standard (0 pg/mL) from the average absorbance of all other readings to obtain corrected absorbance.
- The standard curve is generated by the corrected absorbance of all standard levels on the ordinate against the standard concentration on the abscissa using point-topoint or log-log paper. Appropriate computer assisted data reduction programs may also be used for the calculation of results

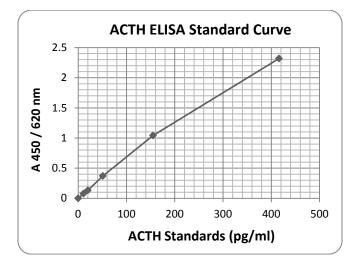
The human ACTH concentrations for the controls and the patient samples are read directly from the standard curve using their respective corrected absorbance.

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#### **EXAMPLE DATA AND STANDARD CURVE**

A typical absorbance data and the resulting standard curve from this EDTA plasma ACTH ELISA are represented. This curve should not be used in lieu of standard curve generated with each assay.

Well I.D.	OD 450/650 nm Absorbance			Results
1.0.	Readings	Average	Corrected	
Std-1: 0 pg/mL	0.025	0.027	0.000	
	0.029			
Std-2: 11 pg/mL	0.107	0.105	0.074	
5.0 = pg=	0.102	*****		
Std-3: 20 pg/mL	0.160	0.160	0.133	
10	0.160			
Std-4: 51 pg/mL	0.400	0.398	0.371	
	0.395		0.0	
Std-5: 155 pg/mL	1.057	1.069	1.042	
ota o. 100 pg/m2	1.081			
Std-6: 416 pg/mL	2.316	2.346	2.319	
ota o. 110 pg/m2	2.375			
Control 1	0.244	0.252	0.225	22.0 ng/ml
22	0.260	1:101	5.420	32.0 pg/mL
Control 2	0.676	0.695	0.668	07.1 ng/ml
	0.714			97.1 pg/mL



### **EXPECTED VALUES**

EDTA plasma samples from normal healthy adults ages 20 – 60 were collected and measured with this ELISA.

The recommended **normal range** for ACTH concentration by using this ELISA is between 1 - 72 pg/mL. We strongly recommend for each clinical laboratory to establish its own normal range by measuring EDTA plasma samples with this ELISA. Please note that sample collection time may have impact on the ACTH normal range.

# LIMITATION OF THE PROCEDURE

- This ACTH assay requires EDTA-plasma sample for testing. Serum sample may show a lower ACTH level and must not be used, because ACTH is not stable in serum.
- Because of 24 hours circadian rhythms of circulating ACTH levels, the time of day the sample was collected should be considered when interpreting test results. Therefore, a normal ACTH test result doesn't rule out related diseases.

- 3. For sample values reading greater than the highest standard, it is recommended to re-assay samples with dilution (i.e. 1:10 or 1:100 with standard zero).
- 4. Water deionized with polyester resins may inactivate the horseradish peroxidase enzyme.

#### **QUALITY CONTROL**

To assure the validity of the results each assay should include adequate controls.

# PERFORMANCE CHARACTERISTICS

#### Sensitivity

The analytical sensitivity (LLOD) of the ACTH ELISA as determined by the 95% confidence limit on 8 replicate determinations of zero standard is less than 1 pg/mL.

#### High Dose "hook" effect

This assay has showed that it did not have any high dose "hook" for ACTH levels up to 10,000 pg/mL.

#### Precision

The intra-assay precision was validated by measuring three control samples with 16 replicate determinations.

Sample #	Mean ACTH Value (pg/mL)	CV (%)
1	36.1	7.6
2	66.5	8.6
3	276.9	10.3

The inter-assay precision was validated by measuring two control levels in duplicate in 16 individual assays.

Sample #	Mean ACTH Value (pg/mL)	CV (%)
1	32.1	7.1
2	261.0	5.3

#### Linearity

Two ACTH standard levels were diluted with standard zero (i.e. level 1 standard) and tested. The results of ACTH percent recovery value in pg/mL are as follows:

DILUTION	OBSERVED VALUE (pg/mL)	RECOVERY %
Neat A	416	-
1:2	231.4	111
1:4	102.1	98
1:8	52.1	100
1:16	26.3	101
Neat B	155	-
1:2	80.3	104
1:4	38.6	100
1:8	20.5	106
1:16	9.3	96

Two EDTA plasma samples were collected, spiked with a high ACTH standard and tested. The results of ACTH percent recovery value in pg/mL are as follows:

DILUTION	OBSERVED VALUE (pg/mL)	RECOVERY %
Neat A	184.1	-
1:2	105.8	115
1:4	58.5	127
1:8	22.6	98
Neat B	33.3	-
1:2	17.1	103
1:4	9.4	113
1:8	5.1	121

#### Spike Recovery

Two EDTA plasma samples and three assay standards (45, 135 and 405 pg/mL) were combined at equal volumes and tested. The results are as follows:

DILUTION	OBSERVED VALUE (pg/mL)	EXPECTED VALUE (pg/mL)	RECOVERY %
Neat A	12.5	=	-
Std-4	25.1	28.8	87
Std-5	73.5	73.8	100
Std-6	254.0	208.8	122
Neat B	21.5	-	-
Std-4	26.1	33.3	79
Std-5	66.8	78.3	85
Std-6	208.1	213.3	98

# **WARRANTY**

This product is warranted to perform as described in its labeling and literature when used in accordance with all instructions. Epitope Diagnostics, Inc. DISCLAIMS ANY IMPLIED WARRANTY OF MERCHANTABILITY OR FITNESS FOR A PARTICULAR PURPOSE, and in no event shall Epitope Diagnostics, Inc. be liable for consequential damages. Replacement of the product or refund of the purchase price is the exclusive remedy for the purchaser. This warranty gives you specific legal rights and you may have other rights, which vary from state to state.

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# TECHNICAL ASSISTANCE AND CUSTOMER SERVICE

For technical assistance or place an order, please contact Epitope Diagnostics, Inc. at (858) 693-7877 or fax to (858) 693-7678. www.epitopediagnostics.com

This product is developed and manufactured by **Epitope Diagnostics, Inc.** 

San Diego, CA 92121, USA





MDSS GmbH Schiffgraben 41 30175 Hannover, Germany

Manufacturer	$\Sigma$ No. of tests	
REF Catalog Number	Keep away from heat and direct sun light	
CONC Concentrate	Store at	
IVD In Vitro Diagnostic Device	Use by	
Read instructions before use	LOT Lot No.	
EC REP Authorized Representative In Europe		

# **ACTH ELISA: Condensed Assay Protocol**

1. 200 µL standards, controls and patient samples

